

TOTAL SYNTHESIS OF *MYO*-INOSITOL-1-PHOSPHATE-4,5-PYROPHOSPHATE,
A NOVEL SECOND MESSENGER ANALOGUE, VIA *MYO*-INOSITOL-1-
PHOSPHATE-4,5-BISPHOSPHOROTHIOATE

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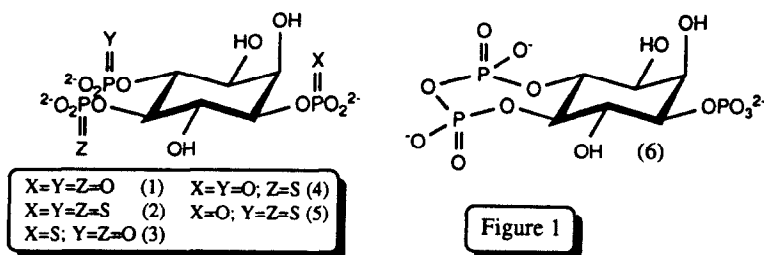
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Abstract: The synthesis of the novel analogues of *myo*-inositol 1,4,5-trisphosphate, *myo*-inositol 1-phosphate 4,5-bisphosphorothioate and *myo*-inositol 1-phosphate 4,5-pyrophosphate is reported; the latter was prepared via desulphurisation and intramolecular coupling.

INTRODUCTION

It is now generally accepted that D-*myo*-inositol 1,4,5-trisphosphate (IP₃) (1) (Fig. 1), released by receptor-mediated phospholipase C-catalysed cleavage of phosphatidylinositol 4,5-bisphosphate, is the second messenger linking the spatially separated events of receptor stimulation and release of intracellular calcium from internal stores^{1,2}. IP₃ is metabolised via two pathways³: deactivation by a 5-phosphatase to 1,4-IP₂ or phosphorylation by a 3-kinase to 1,3,4,5-IP₄. The function of the latter still remains controversial and IP₄ may gate a plasma membrane Ca²⁺ channel⁴. IP₃ acts through an intracellular receptor which has been isolated⁵, cloned and sequenced^{6,7} and reconstituted⁸.

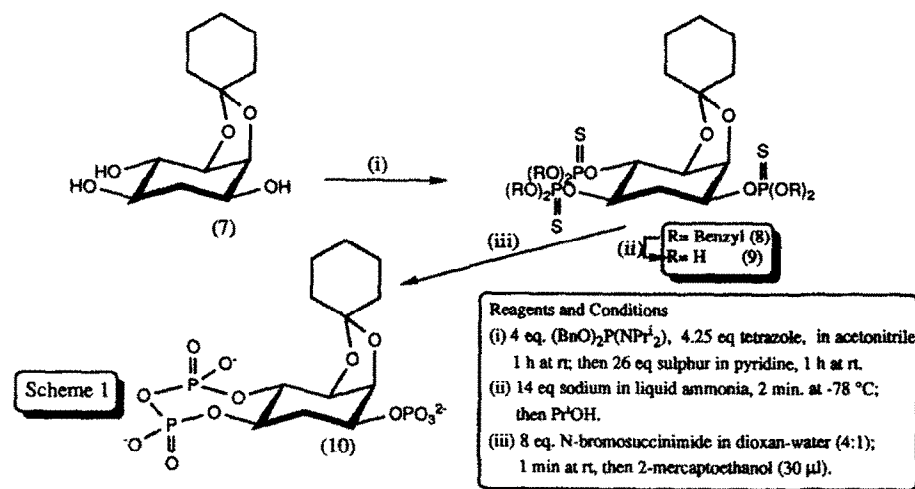
We have sought to develop synthetic routes to inositol phosphates⁹ and especially to prepare non-hydrolysable analogues such as phosphorothioates^{9,10,11}. Our synthesis of *myo*-inositol 1,4,5-trisphosphorothioate (IPS₃) (2)¹² (Fig. 1) has provided an analogue that is a potent releaser of calcium¹³⁻¹⁵ and yet is resistant to phosphatase-catalysed deactivation¹⁶. Other biologically potent Ca²⁺-mobilising synthetic phosphorothioate analogues include *myo*-inositol 1-phosphorothioate 4,5-bisphosphate (3)¹⁷ and *myo*-inositol 1,4-bisphosphate 5-phosphorothioate (4)^{18,19}. It is clear that such analogues offer considerable potential for investigation and modification of the complex metabolism of IP₃ and this has been recognized by other groups^{20,21}.



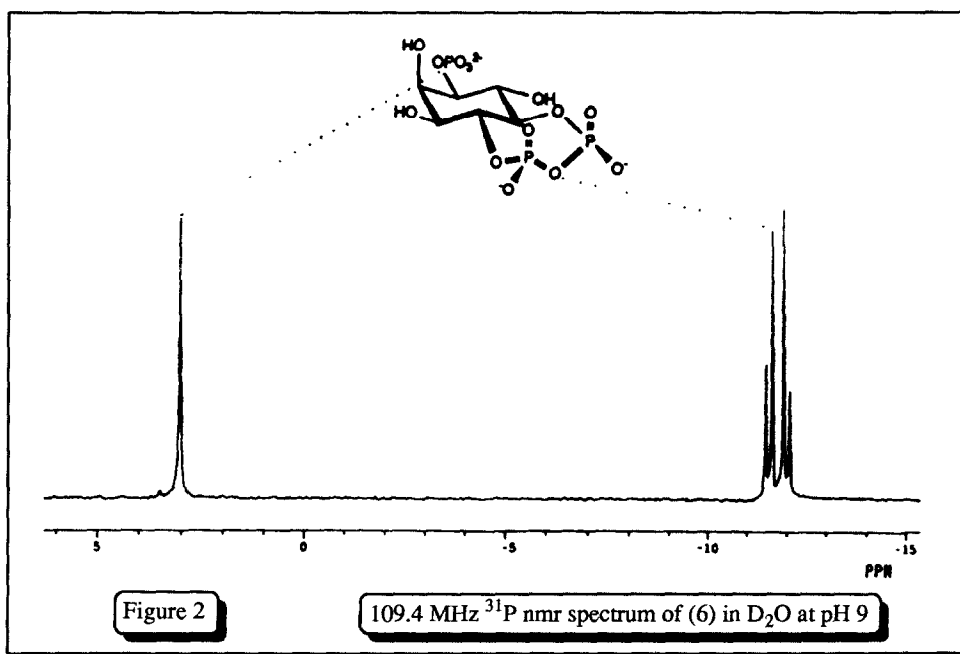
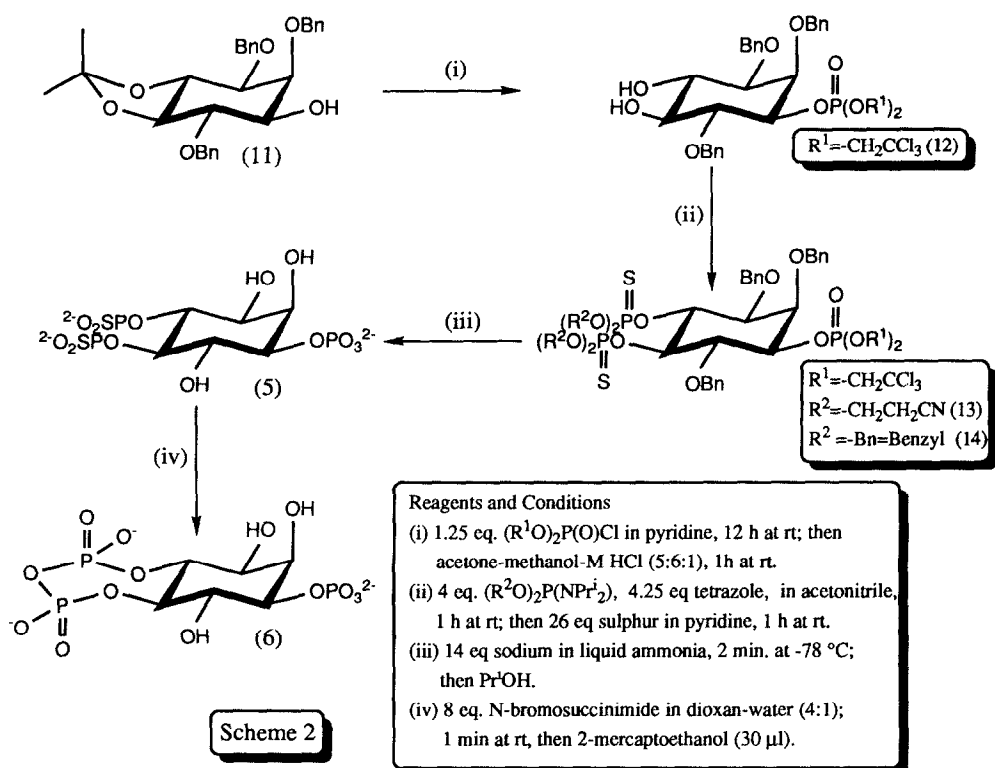
In structure-activity studies, the vicinal 4,5-bisphosphate moiety of IP_3 has been identified as being crucial for intracellular Ca^{2+} release^{3,9-11}. Clearly, chemical modification at this locus may be one of the keys for designing potential IP_3 receptor antagonists. An obvious synthetically-challenging target is the pyrophosphate (6) where the 4,5-phosphate groups have been linked together. As well as comprising a novel modification to the IP_3 structure, formation of the pyrophosphate produces a less polar analogue that can potentially be converted to IP_3 enzymatically in cells. We have previously reported a new route to such seven-membered pyrophosphates by desulphurisation of a vicinal bisphosphorothioate using *N*-bromosuccinimide (NBS)²². We report here the total synthesis of (6) via the novel phosphorothioate analogue *myo*-inositol 1-phosphate 4,5-bisphosphorothioate (5).

Initial experiments on the feasibility of using polyphosphorothioate derivatives of inositol to prepare cyclic pyrophosphates by our desulphurisation method²² were performed on (7), a protected precursor of D-6-deoxy-*myo*-inositol 1,4,5-trisphosphate²³. Thus, D-2,3-*O*-cyclohexylidene-6-deoxy-*myo*-inositol²⁴ (7) (Scheme 1) was converted into its trisphosphorothioate ester (8) using bisbenzyl-diisopropylaminophosphine-tetrazole²⁵, followed by oxidation of the resulting trisphosphite using sulphur in pyridine¹². Deprotection using sodium in liquid ammonia²⁶ removed the benzyl protecting groups and purification of the crude product on DEAE Sepharose using a gradient of triethylammonium bicarbonate (TEAB) gave D-2,3-*O*-cyclohexylidene-6-deoxy-*myo*-inositol-1,4,5-trisphosphorothioate (9) as its triethylammonium salt. This compound has the advantage that on NBS-mediated desulphurisation there are no neighbouring vicinal hydroxyl groups to facilitate intramolecular cyclisation to form cyclic 5-membered phosphates. Furthermore, we expected that conformational restriction of the cyclohexane

ring, due to the presence of the 2,3-ketal might aid the ring closure of the *trans*-vicinal 4,5-phosphates. Consequently, the 1-phosphorothioate group should be desulphurised to the corresponding phosphate and we expected intramolecular coupling of the activated 4,5-bisphosphorothioate to form the cyclic pyrophosphate. Indeed, desulphurisation of (9) using NBS gave D-2,3-*O*-cyclohexylidene-6-deoxy-*myo*-inositol-1-phosphate-4,5-cyclic pyrophosphate (10) in > 90% yield, demonstrating the feasibility of our approach in the inositol series. (10) Was purified by ion-exchange chromatography. ^{31}P nmr spectroscopy of (10) (109.4 MHz, D_2O pH9, ext. H_3PO_4 ref.) showed clearly a peak at δ -1.84 ppm assignable to the 1-phosphate group and an AB system with δ -12.52 and -13.02 ppm, $^2J_{\text{PP}}$ 16.8 Hz, assignable to the 4,5-pyrophosphate system.



To avoid complications due to potential cyclisation of an activated 1-phosphorothioate group during desulphurisation of IPS_3 (2) we have designed a synthesis of *myo*-inositol 1-phosphate 4,5-bisphosphorothioate (5) as a precursor of (6). (\pm) -2,3,6-tri-*O*-benzyl-1[bis(2,2,2-trichloroethyl)phospho]-*myo*-inositol (12) was synthesised as described^{19,27} from the protected mono-alcohol (11)²⁸. Bisphosphitylation of (12) with either bis(2-cyanoethyl)diisopropylaminophosphine²⁹ or bisbenzylidiisopropylaminophosphine²⁵ gave the corresponding 4,5-bisphosphites, which were oxidised to the respective 4,5-bisphosphorothioates (13) and (14) with sulphur in pyridine (Scheme 2). Deprotection by



sodium in liquid ammonia as above gave crude DL-*myo*-inositol-1-phosphate-4,5-bisphosphorothioate (5) which was purified by ion-exchange chromatography on either DEAE Sephadex A-25 or Q-Sepharose to give the pure triethylammonium salt of (5), quantified by Briggs phosphate assay³⁰, in 83% yield.

DL-(5) Was a potent agonist for intracellular Ca^{2+} mobilisation in permeabilised SH-SY5Y neuroblastoma cells³¹. It mobilised Ca^{2+} with a potency slightly less than IP_3 and similar to IPS_3 (2). Having a 5-phosphorothioate group, (5) is resistant to degradation by IP_3 -5-phosphatase and stimulated a persistent release of Ca^{2+} like IPS_3 ^{3,31}. It inhibited IP_3 -5-phosphatase potently with a K_i of $1.3 \pm 0.3 \mu\text{M}$ similar to IPS_3 ³² suggesting that D-(5) would have a submicromolar K_i for this enzyme. Full biological details will be published elsewhere.

(5) Was desulphurised with NBS and gave the crude pyrophosphate (6) in high yield with some evidence of phosphate migration (24%) and straight desulphurisation to IP_3 (7%). (6) Was purified by anion exchange chromatography on Q-Sepharose as above to give the triethylammonium salt (67% yield). It exhibited a ^{31}P n.m.r spectrum (Figure 2) showing clearly the presence of the 1-phosphate (δ 3.04 ppm) and the 4,5-pyrophosphate [δ -11.61 (pos. 4) and -12.02 ppm (pos. 5), $^2J_{\text{pp}}$ 16.8 Hz], the latter resonating as the expected AB system. Biological evaluation of (6) is in progress.

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